

Ainhoa Lapitz¹, Mikel Azkargorta,^{2,3} Piotr Milkiewicz,^{4,5} Paula Olaizola,^{1,3} Ekaterina Zhuravleva,⁶ Marit M. Grimsrud,^{7,8} Christoph Schramm,^{9,10,11} Ander Arbelaiz,¹ Colm J. O'Rourke,⁶ Adelaida La Casta,¹ Malgorzata Milkiewicz,¹² Tania Pastor,¹ Mette Vesterhus,^{7,13} Raul Jimenez-Agüero,¹ Michael T. Dill,^{14,15} Angela Lamarca,¹⁶ Juan W. Valle,¹⁶ Rocio I.R. Macias,^{3,17} Laura Izquierdo-Sanchez,^{1,3} Ylenia Pérez Castaño,^{1,18} Francisco Javier Caballero-Camino,¹ Ioana Riano,^{1,19} Marcin Krawczyk,²⁰ Cesar Ibarra,²¹ Javier Bustamante,²¹ Luiz M. Nova-Camacho,²² Juan M. Falcon-Perez,^{3,23,24} Felix Elortza,^{2,3} Maria J. Perugorria,^{1,3,25} Jesper B. Andersen,⁶ Luis Bujanda,^{1,3,25} Tom H. Karlsen,⁷ Trine Folseraas,^{7,26} **Pedro M. Rodrigues**,^{1,3,24,27} **Jesus M. Banales**^{1,3,24,27}

¹Department of Liver and Gastrointestinal Diseases, Biodonostia Health Research Institute – Donostia University Hospital, University of the Basque Country (UPV/EHU), 20014, San Sebastian, Spain; ²Proteomics Platform, CIC bioGUNE, Basque Research and Technology Alliance (BRTA), ProteoRed-ISCIII, Bizkaia Science and Technology Park, 48160, Derio, Spain; ³National Institute for the Study of Liver and Gastrointestinal Diseases (CIBERehd), ISCIII, 28220, Madrid, Spain; ⁴Liver and Internal Medicine Unit, Department of General, Transplant and Liver Surgery, Medical University of Warsaw, 02-097, Warsaw, Poland; ⁵Translational Medicine Group, Pomeranian Medical University, 70-204, Szczecin, Poland; ⁶Biotech Research and Innovation Centre, Department of Health and Medical Sciences, University of Copenhagen, 2200, Copenhagen, Denmark; ⁷Norwegian PSC Research Center, Department of Transplantation Medicine, Division of Surgery, Inflammatory Medicine and Transplantation, Oslo University Hospital, Rikshospitalet, Oslo, Norway; ⁸Institute of Clinical Medicine, University of Oslo, Oslo, Norway; ⁹European Reference Network Hepatological Diseases (ERN RARE-LIVER), Hamburg, Germany; ¹⁰1st Department of Internal Medicine, University Medical Center Hamburg-Eppendorf, Hamburg, Germany; ¹¹Martin Zeitz Centre for Rare Diseases, University Medical Centre Hamburg-Eppendorf, Hamburg, Germany; ¹²Department of Medical Biology, Pomeranian Medical University in Szczecin, 70-111 Szczecin, Poland; ¹³Department of Clinical Science, University of Bergen, Bergen, Norway; ¹⁴Department of Gastroenterology, Infectious Diseases and Intoxication, Heidelberg University Hospital, 69120, Heidelberg, Germany; ¹⁵Experimental Hepatology, Inflammation and Cancer, German Cancer Research Center (DKFZ), 69120, Heidelberg, Germany; ¹⁶Department of Medical Oncology, The Christie NHS Foundation Trust/Division of Cancer Sciences, University of Manchester, Manchester, UK; ¹⁷Experimental Hepatology and Drug Targeting (HEVPHARM), University of Salamanca, Biomedical Research Institute of Salamanca (IBSAL), 37007, Salamanca, Spain; ¹⁸Osakidetza Basque Health Service, Bidasoa IHO, Bidasoa Hospital, Department of Digestive System, Irun, Spain; ¹⁹Clinical Research Unit, Spanish Clinical Research Network (SCRn) - ISCIII, Biodonostia Health Research Institute, San Sebastián, Spain; ²⁰Department of Medicine II, Saarland University Medical Centre, Saarland University, Homburg, Germany; ²¹Osakidetza Basque Health Service, Euzkarraldeko-Enfermeria-Cruces IHO, Cruces University Hospital, Barakaldo, Spain; ²²Osakidetza Basque Health Service, Donostialdea IHO, Donostia University Hospital, Department of Pathology, San Sebastián, Spain; ²³Center for Cooperative Research in Biosciences (CIC bioGUNE), Basque Research and Technology Alliance (BRTA), Exosomes Laboratory, 48160, Derio, Spain; ²⁴Kerbasque, Basque Foundation for Science, 48013, Bilbao, Spain; ²⁵Department of Medicine, Faculty of Medicine and Nursing, University of the Basque Country, UPV/EHU, Leioa, Spain; ²⁶Section of Gastroenterology, Department of Transplantation Medicine, Oslo University Hospital, Oslo, Norway; ²⁷Department of Biochemistry and Genetics, School of Sciences, University of Navarra, Pamplona, Spain.

INTRODUCTION

- Cholangiocarcinoma (CCA)** includes a heterogeneous group of malignancies with dismal prognosis.
- The etiology is unknown but pathologies such as **primary sclerosing cholangitis (PSC)** increase the odds of CCA development (up to 20% life-time risk).
- There is an urgent need of accurate **non-invasive biomarkers** for the early diagnosis of CCA, particularly in high-risk populations.
- Extracellular vesicles (EVs), small membranous spheres found in biofluids, have recently emerged as a potential source of biomarkers.

AIM

Identify new accurate **non-invasive protein biomarkers** in serum EVs to **predict CCA development**, to **early diagnose CCA**, as well as to **estimate prognosis** of patients with CCA.

Understand the potential **origin of serum EV biomarkers** for their liquid biopsy application.

METHODS

- Isolation of serum EVs** from patients with isolated PSC (n=45), PSC without clinical evidences of malignancy at sampling who developed CCA overtime (PSC to CCA; n=25), concomitant PSC-CCA (n=42), CCAs from non-PSC etiology (n=56), hepatocellular carcinoma (HCC, n=34) and healthy individuals (n=55).
- EV characterization by transmission electron microscopy (TEM), immunoblot and nanoparticle tracking analysis (NTA).
- Proteomic analysis of EVs by mass spectrometry (MS).
- Evaluation of the diagnostic efficacy of proteins by receiver operating characteristic (ROC) curves.
- Validation of the diagnostic capacity of biomarkers in total serum by enzyme-linked immunosorbent assay (ELISA)
- Expression analysis of biomarker candidates in **human multi-organ transcriptomes**, in single-cell RNA-sequencing (scRNA-seq) of healthy livers and in scRNA-seq of CCA tumors.
- Evaluation of the use of serum EV protein levels as **survival predictors**.

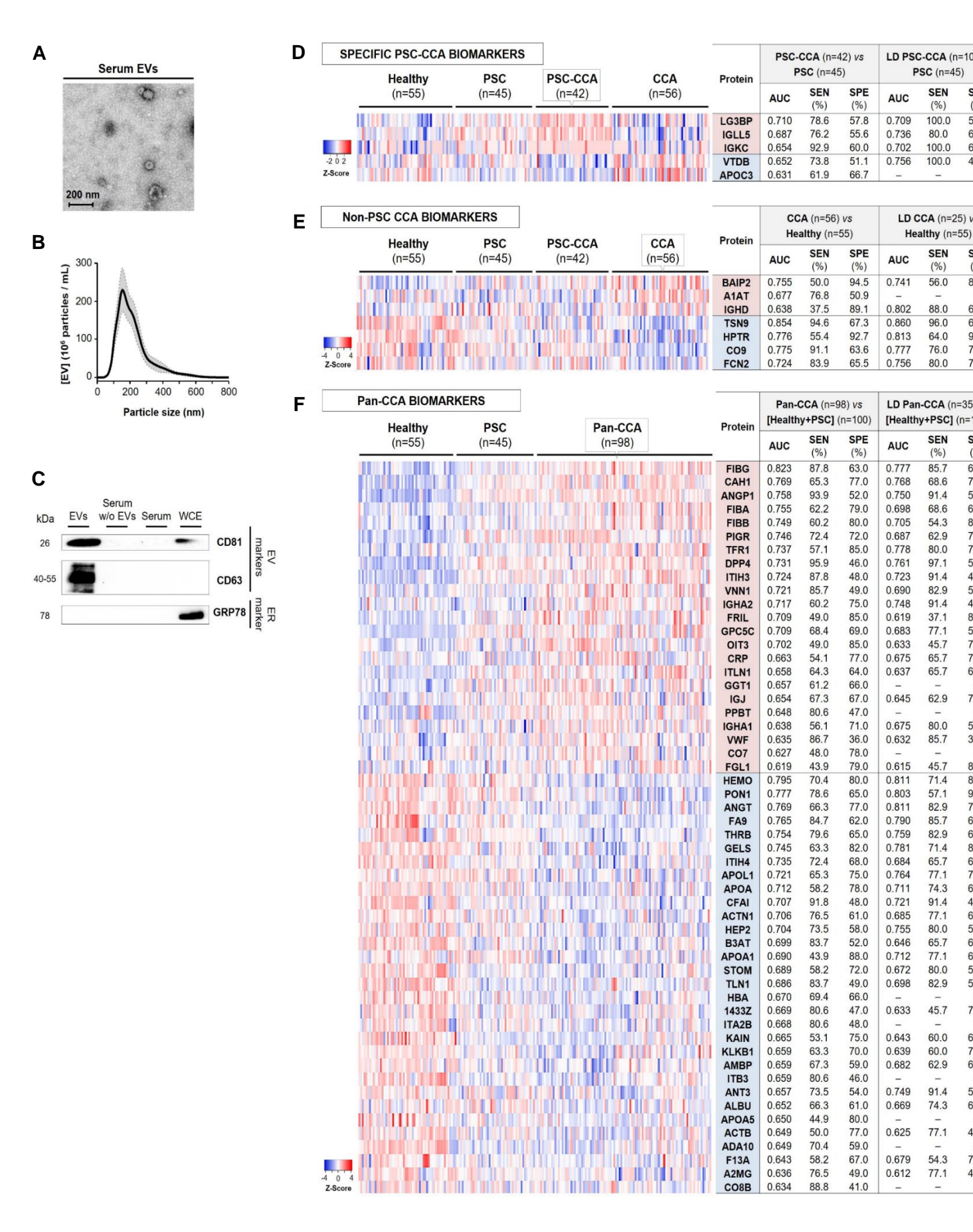
RESULTS

Table 1. Demographic and clinical features of the study cohort (analyzed by MS)

GENERAL DEMOGRAPHICS	Healthy		PSC		PSC to CCA		PSC-CCA		CCA		HCC	
	Obtbpap (n=19)	ImuT0F (n=55)	Obtbpap (n=18)	ImuT0F (n=45)	ImuT0F (n=25)	Obtbpap (n=14)	ImuT0F (n=42)	Obtbpap (n=56)	ImuT0F (n=56)	Obtbpap (n=34)	ImuT0F (n=34)	
Age	mean ± SD (range)	60 ± 6 (57 to 10)	35 ± 13 (18 to 55)	38 ± 13 (18 to 55)	48 ± 14 (18 to 74)	53 ± 12 (17 to 81)	49 ± 14 (18 to 74)	66 ± 10 (48 to 81)	63 ± 9 (48 to 81)			
Sex	Male, n (%)	5 (26.3)	22 (40.0)	15 (83.3)	34 (75.6)	21 (84.0)	19 (71.4)	28 (66.7)	40 (71.4)			
CLINICAL PARAMETERS, n (%)	IBD											
	UC		2 (11.1)	6 (13.3)	6 (34.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	
	Colitis		10 (55.6)	39 (66.7)	16 (84.0)	13 (62.9)	28 (66.7)	11 (8.3)	11 (8.3)	0 (0.0)	0 (0.0)	
	Unspecified		3 (16.7)	4 (8.9)	3 (12.0)	0 (0.0)	1 (11.9)	1 (1.8)	0 (0.0)	0 (0.0)	0 (0.0)	
	Yes		3 (16.7)	5 (11.3)	0 (0.0)	1 (7.1)	1 (2.4)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	
	No		0 (0.0)	0 (0.0)	9 (35.0)	1 (7.1)	4 (9.5)	4 (7.1)	24 (70.6)			
	Cirrhosis		15 (83.3)	43 (85.6)	16 (84.0)	12 (57.1)	38 (90.5)	59 (89.3)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
	Yes		3 (16.7)	2 (4.4)	0 (0.0)	1 (7.1)	0 (0.0)	2 (3.6)	2 (3.6)	0 (0.0)	0 (0.0)	0 (0.0)
	No		0 (0.0)	0 (0.0)	1 (4.0)	0 (0.0)	2 (4.8)	3 (5.4)	20 (58.8)			
	HBV / HCV		18 (100)	48 (100)	24 (86.9)	14 (100)	39 (95.2)	59 (84.4)	14 (41.2)	14 (41.2)		
	Yes		0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (2.4)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
	No		0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
Cholelithiasis		18 (100)	48 (100)	28 (100)	14 (100)	42 (100)	56 (100)	34 (100)	34 (100)			
Yes		0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	
No		0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	
Liver fluke		18 (100)	48 (100)	28 (100)	14 (100)	42 (100)	56 (100)	34 (100)	34 (100)			
Yes		0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	
No		0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	
CCA subtype												
iCCA					15 (60.0)	7 (50.0)	19 (45.2)	18 (32.1)				
pCCA					2 (8.0)	2 (8.0)	2 (4.8)	1 (1.8)				
dCCA					7 (28.0)	2 (8.0)	10 (23.8)	28 (44.6)				
Disease status												
LAD					11 (44.0)	7 (50.0)	15 (35.7)	11 (19.6)				
MD					2 (8.0)	2 (8.0)	2 (4.8)	2 (3.6)				
No					4 (16.0)	5 (35.7)	16 (36.1)	37 (66.1)				
CCA-related surgery												
Liver transplant					18 (72.0)	6 (42.9)	11 (25.0)	0 (0.0)				
Tumor resection					2 (8.0)	3 (21.4)	17 (38.1)	19 (33.9)				
Esophagectomy					1 (4.0)	0 (0.0)	1 (2.4)	0 (0.0)				
BIOCHEMICAL PARAMETERS	ALT (U/L)	mean ± SD	108 ± 55	91 ± 65	105 ± 77	129 ± 209	121 ± 164	88 ± 123	52 ± 81			
	NAK, n (%)		1 (7.1)	2 (4.0)								
	AST (U/L)	mean ± SD	69 ± 63	65 ± 49	92 ± 61	142 ± 209	103 ± 120	67 ± 84	48 ± 50			
	NAK, n (%)		1 (7.1)	1 (2.0)								
	GOT (U/L)	mean ± SD	301 ± 224	281 ± 156	286 ± 247	590 ± 240	309 ± 230	601 ± 59	378 ± 247			
	NAK, n (%)		1 (7.1)	1 (2.0)								
	Total bilirubin (mg/dL)	mean ± SD	1.8 ± 0.9	1.2 ± 1.3	5.2 ± 8.4	9.4 ± 9.4	6.2 ± 7.4	3.8 ± 5.2	6.6 ± 4.0			
	NAK, n (%)		2 (8.0)	2 (4.0)								
	ALP (U/L)	mean ± SD	228 ± 161	238 ± 172	382 ± 257	381 ± 340	425 ± 256	323 ± 279	1167 ± 194			
	NAK, n (%)		1 (7.1)	1 (2.0)								
	CA19-9 (U/mL)	mean ± SD	51.5 ± 34	27.6 ± 36	167 ± 103	185 ± 132	208 ± 120	207 ± 103	219 ± 153			
	NAK, n (%)		3 (16.7)	4 (8.9)	0 (0.0)	1 (7.1)	2 (4.8)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	
AFP (ng/mL)	mean ± SD	4.7 ± 4.4	3.9 ± 3.5	4.0 ± 4.2	12.9 ± 21.0	8.1 ± 15.4	14.4 ± 37.3	5.5 ± 5.8				
NAK, n (%)		0 (0.0)	27 (80.0)	2 (8.0)	2 (14.3)	9 (21.4)	34 (80.7)	0 (0.0)	0 (0.0)	0 (0.0)		

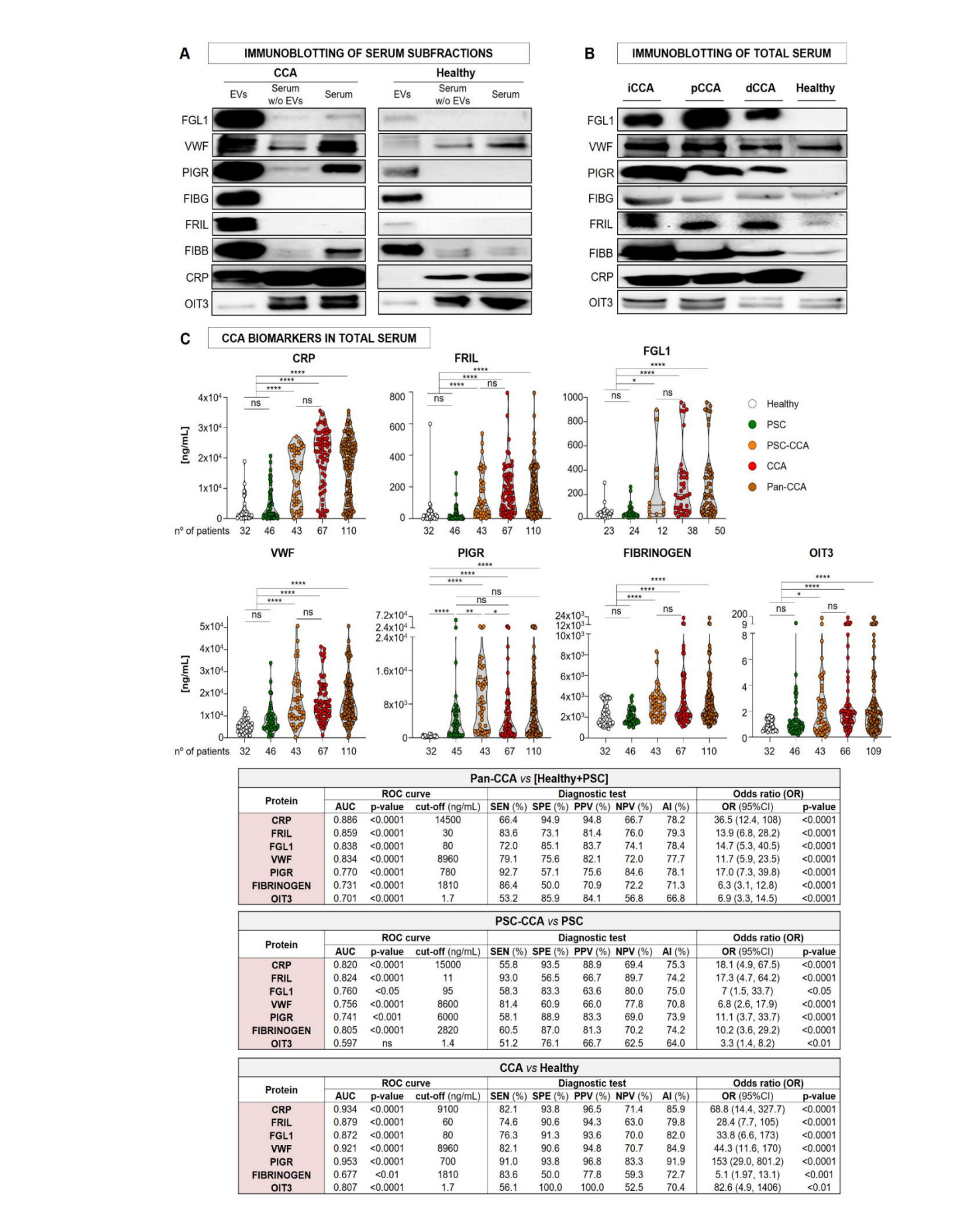
* Healthy (1855 samples previously analyzed in the Obtbpap MS), PSC (1846 samples analyzed also in the Obtbpap MS), PSC-CCA (1242 samples analyzed in the Obtbpap MS).

Figure 1. Serum EV-protein biomarkers for CCA diagnosis according to tumor etiology



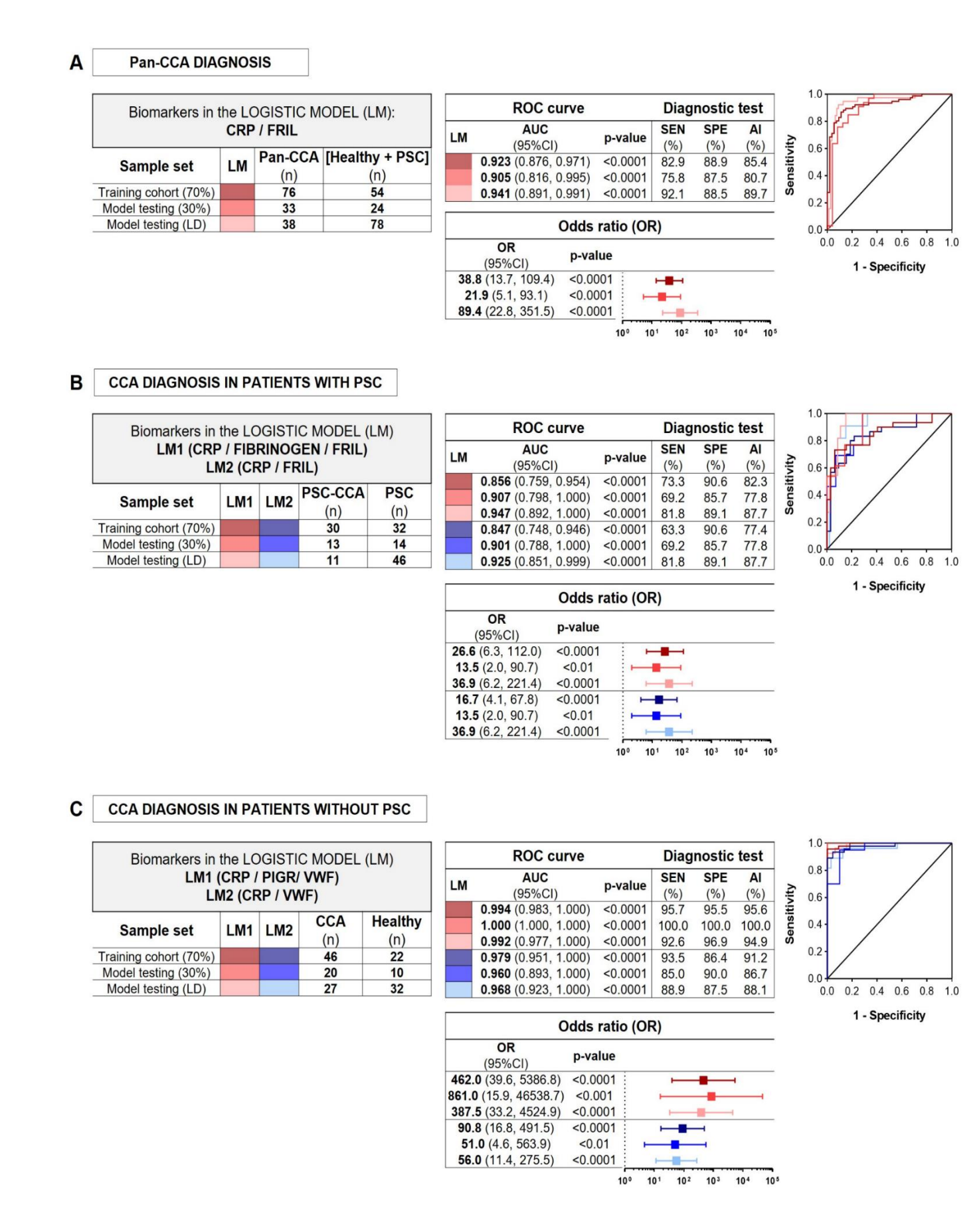
Characterization of serum EVs by (A) TEM, (B) NTA and (C) immunoblotting. Biomarkers for the specific diagnosis of (D) CCA in patients with PSC, (E) CCA in patients with PSC and (F) CCA regardless etiology (Pan-CCA biomarkers). Enriched proteins are colored in red and proteins with lower abundance in blue.

Figure 2. EV-protein biomarkers are detected using total serum and aid the diagnosis of CCA



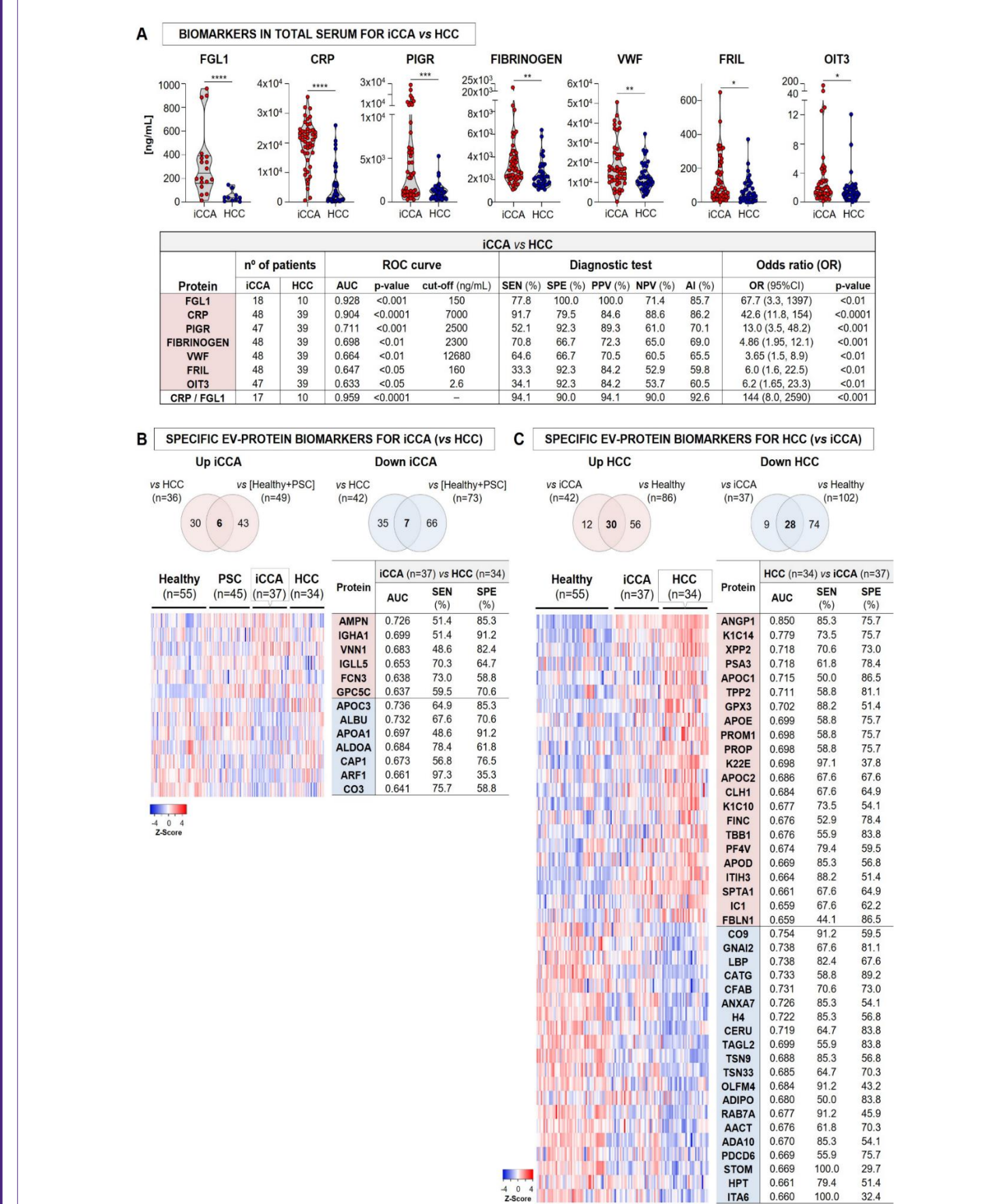
(A) Immunoblots of selected biomarkers in serum subfractions of patients with CCA and healthy individuals. (B) Immunoblots of biomarkers in total serum of patients with iCCA, pCCA or dCCA and healthy individuals. (C) Biomarker levels measured by ELISA in serum samples from patients with PSC, PSC-CCA, non-PSC CCA and healthy individuals and their individual diagnostic values.

Figure 3. Logistic models combining ELISA-validated serum protein biomarkers for accurate CCA diagnosis



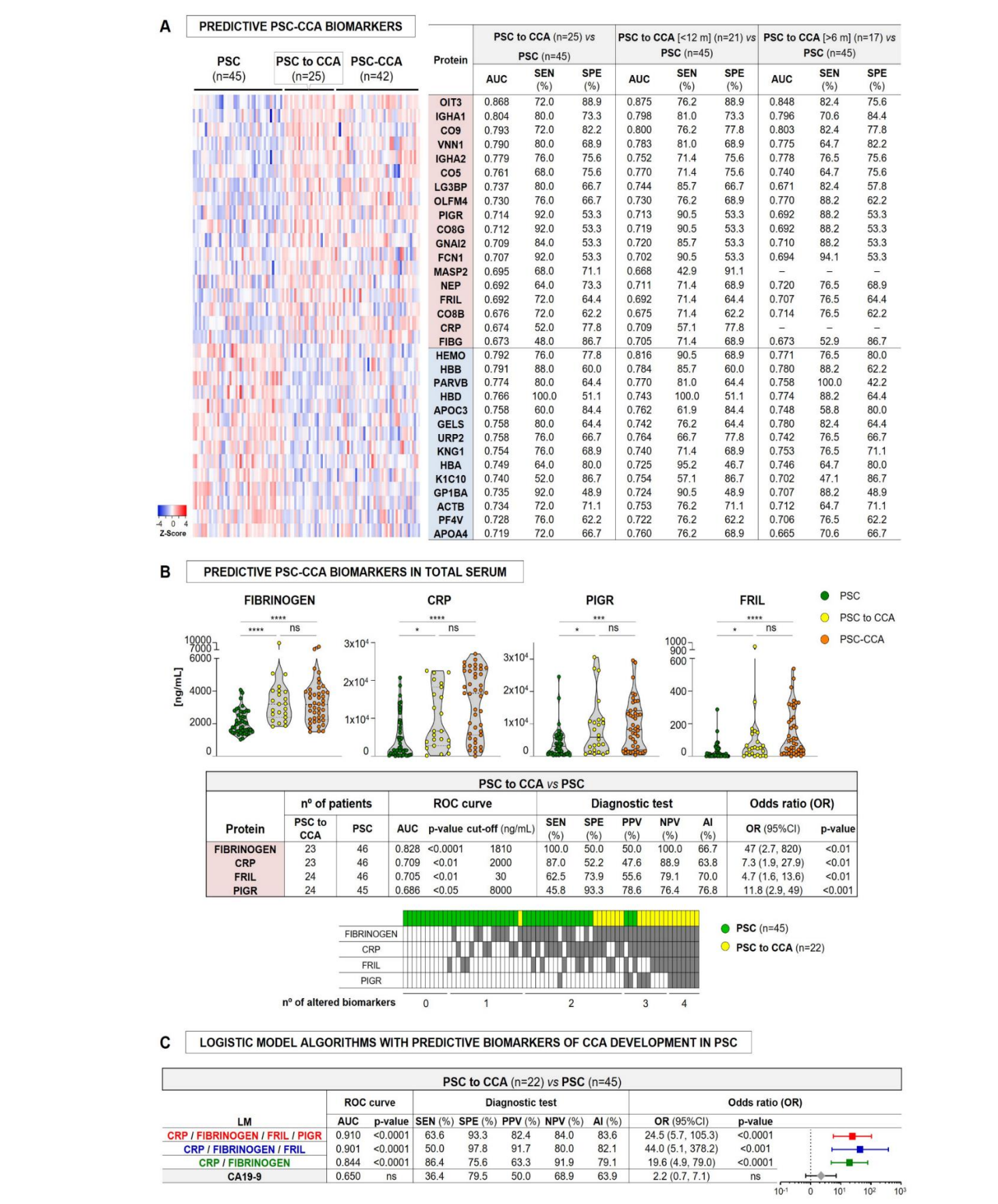
Logistic models combining ELISA-validated serum protein biomarkers enables the accurate diagnosis of CCA in patients with or without PSC. Binary logistic regression models in the training (70%), as well as in testing 30% and local disease (LD) cohorts for CCA diagnosis (A) regardless disease etiology, (B) in patients with PSC and (C) in patients without PSC.

Figure 4. Serum EV-protein biomarkers for the differential diagnosis of iCCA vs HCC



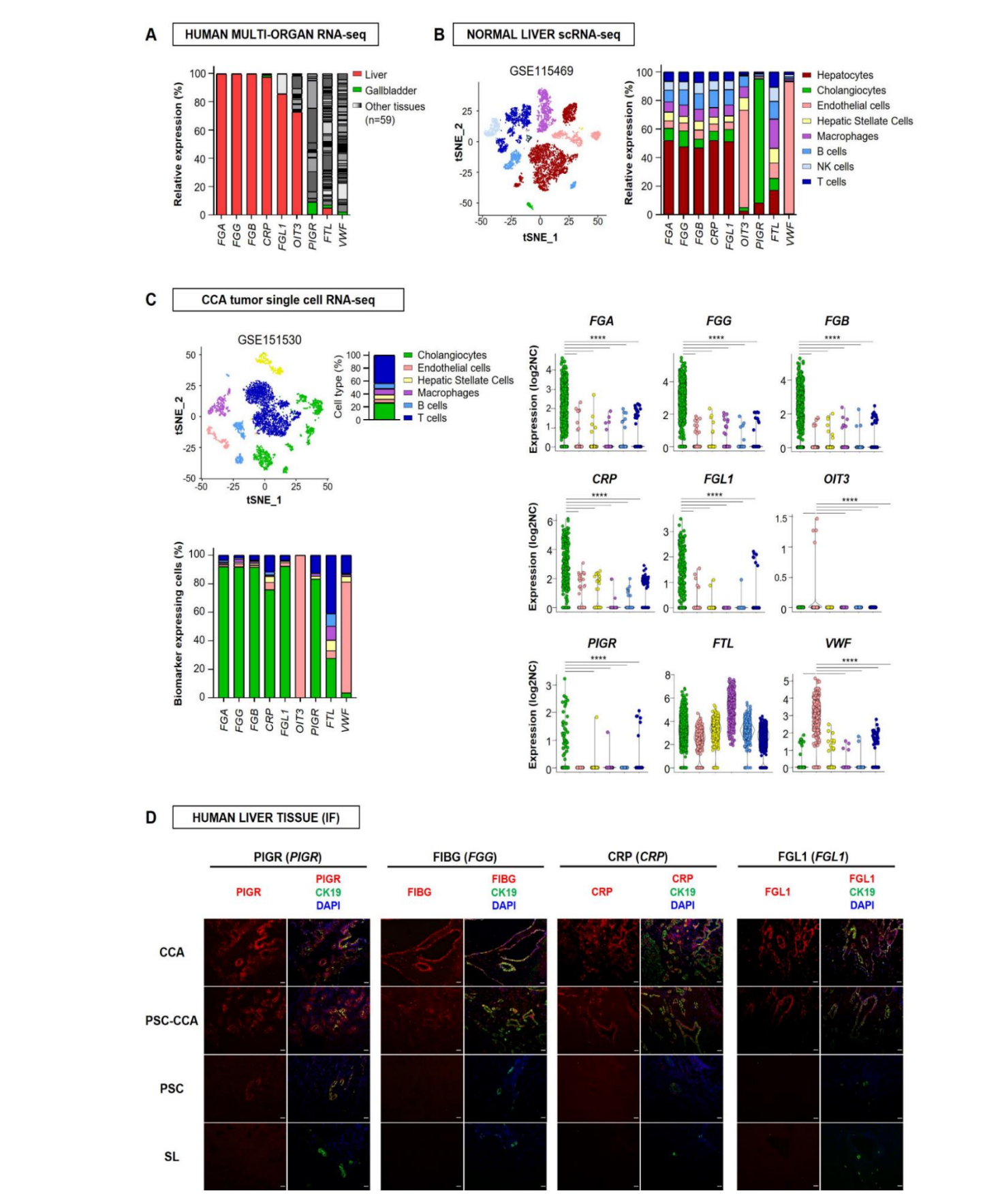
(A) Levels and diagnostic values of FGL1, CRP, PIGR, FIBRINOGEN, VWF, FRIL and OIT3 measured by ELISA in serum samples from patients with iCCA and HCC. Heatmaps, Venn diagrams and diagnostic values of specific EV-proteins for the diagnosis of (B) iCCA and (C) HCC.

Figure 5. Serum proteins allow the prediction of CCA development in patients with PSC



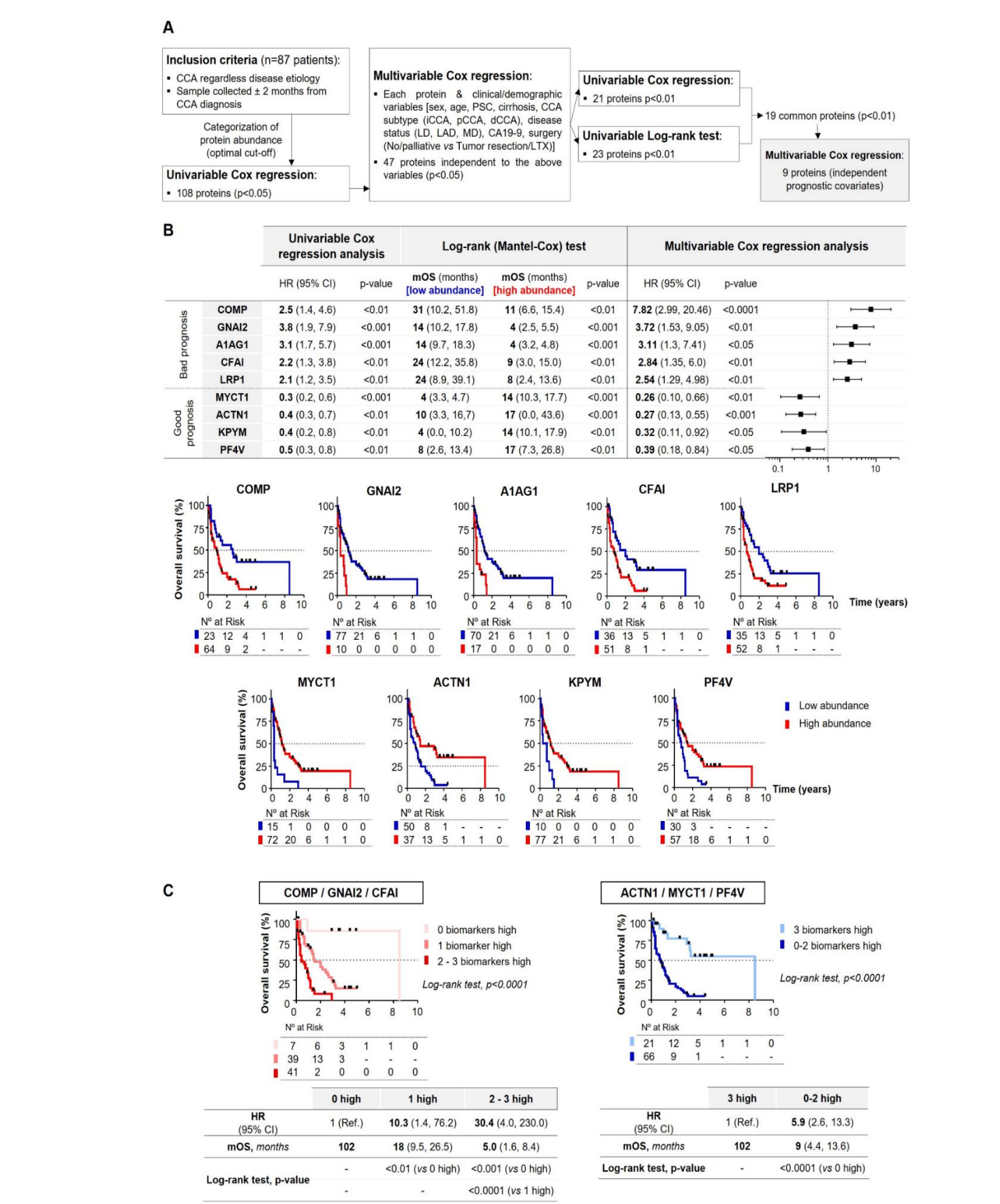
(A) Specific EV-proteins for the differential identification of patients with PSC who progressed to CCA over-time (PSC to CCA) and non-malignant PSC. (B) Levels and diagnostic values of FIBRINOGEN, CRP, PIGR and FRIL in total serum from PSC to CCA, PSC-CCA patients and non-malignant PSC. (C) Binary logistic regression models for the prediction of CCA development in patients with PSC.

Figure 6. Potential origin of serum EV-protein biomarkers



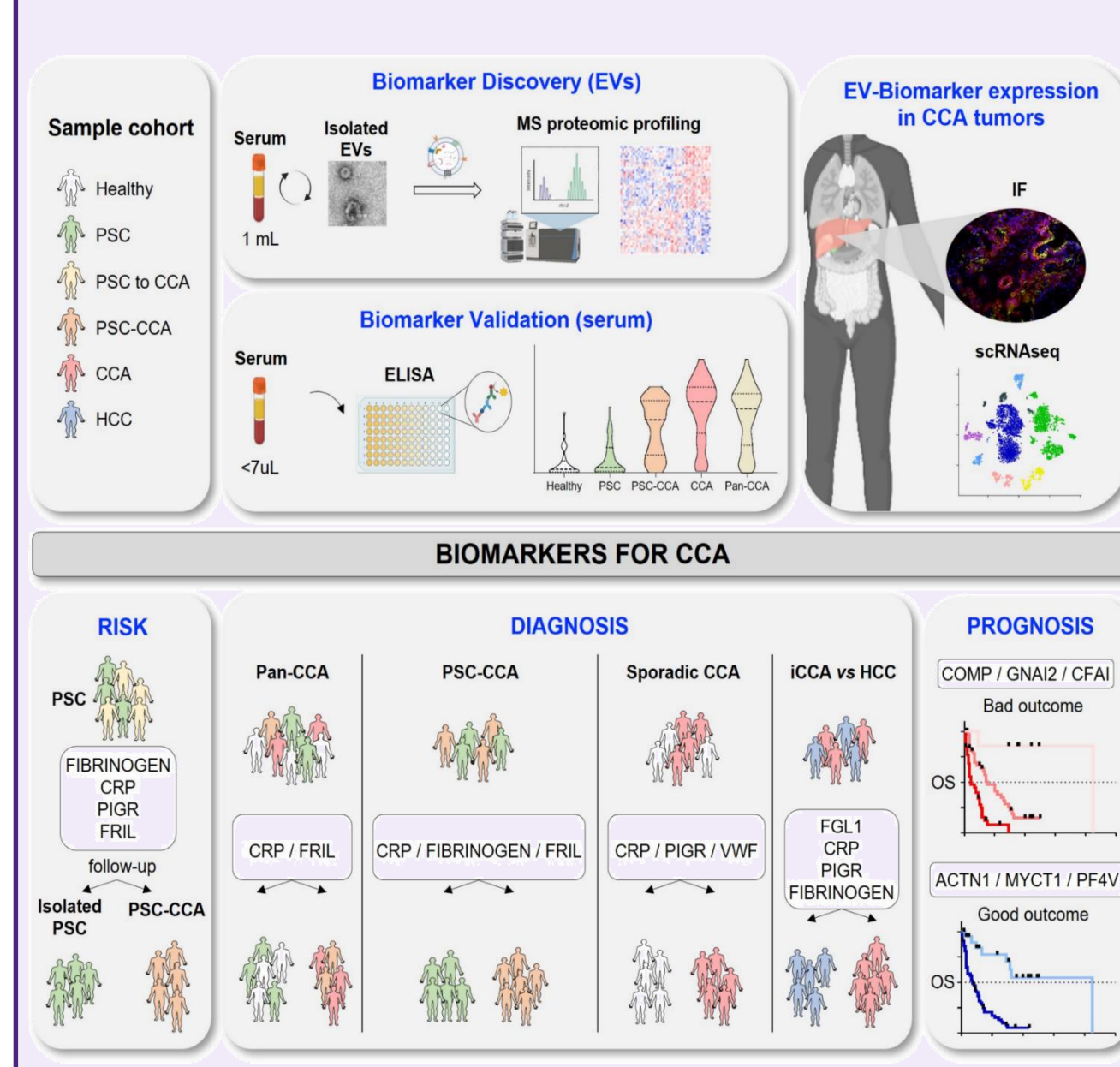
(A) Expression of candidate serum biomarkers in 61 human tissues/organs from the Human Protein Atlas. (B) tSNE plot and expression of candidate biomarkers in each liver cell type from normal liver scRNA-seq (GSE115469). (C) tSNE plot and cell type proportion from iCCA tumors (GSE151530). Biomarker expressing-positive cells and relative expression within iCCA tumor cells. (D) Immunofluorescence images of biomarkers and co-localization with CK19+ positive cells.

Figure 7. Association of serum EV-protein levels with patients' outcome



(A) Strategy used to define prognostic biomarkers. (B) Multivariable analysis of serum EV-proteins with independent prognostic value to clinical variables. (C) Kaplan Meier curve, Cox regression analysis and Log-rank test of patients with CCA according to the "bad prognostic" (COMP/GNA2/CFAI) and "good prognostic" (ACTN1/MYCT1/PFV) panels.

CONCLUSIONS



- Serum EVs contain protein biomarkers for:**
- the prediction of CCA development in PSC
 - the early tumor detection in individuals with PSC, in individuals without PSC and also for CCAs regardless of disease etiology
 - the differential diagnosis between iCCA and HCC
 - the prognostic estimation of individuals with CCA

Serum EV biomarkers are amenable to be detected using total serum.

Most of these candidate biomarkers are preferentially expressed in malignant cholangiocytes within CCA tumors, representing a novel tumor cell-derived liquid biopsy for personalized medicine.

DISCLAIMER

The findings presented in this abstract have been recently published as **Liquid biopsy-based protein biomarkers for risk prediction, early diagnosis and prognostication of cholangiocarcinoma in Journal of Hepatology** [2023 Mar 1; S0168-8278(23)00159-9. doi: 10.1016/j.jhep.2023.02.027]. We acknowledge and thank the journal for allowing us to present this work at the AMMF 2023 European Cholangiocarcinoma Conference.

CONTACT INFO

pedro.rodrigues@biodonostia.org
jesus.banales@biodonostia.org